Exploiting the synthetic capacity of microbes for the production of novel value-added biochemicals

Kristala Jones Prather

Theodore T. Miller Career Development Associate Professor Department of Chemical Engineering Massachusetts Institute of Technology

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Microbes as Chemical Factories



Improvement of natural producers

Microbes as Chemical Factories

- Antibiotics/Antimicrobials
- Other therapeutics (lovastatin)
- Amino Acids
- Organic Acids



Re-constitution of natural pathways in heterologous hosts

Changing the paradigm^{*} – from " CH_2 " to " CH_2O "



*Prof. Bradley D. Olsen, MIT Chemical Engineering

"Retro-biosynthetic" Pathway Design*

- Integration of Biocatalysis ("Parts" selection) and Metabolic Engineering ("Systems" assembly, analysis)
- (Others) On-going work on algorithms for biosynthetic pathway design
- Elucidation of Design Principles
- Development of Design and Assembly Tools ("Devices")

* Curr. Opin. Biotechnol. 2008. 19:468-474

Candidates for Target Molecules

Building Blocks

C	1,4-diacids (succinic, fumaric and malic)				
	2,5-furan dicarboxylic acid				
_	3-hydroxypropionic acid				
Т	aspartic acid				
	Glucaric Acid				
	glutamic acid				
	itaconic acid	MARKED BAS			
	levulinic acid				
	3-Hydroxybutyrolactone				
	glycerol				
	sorbitol				
	xylitol/arabinitol				

August 2004

Glucaric Acid



- Found in fruits and vegetables, mammals
- No known microbial pathway
- Previously studied for cholesterol-reducing, chemotherapeutic effects
- Potential use as building block for polymeric materials (nylons), detergents
- Produced chemically through acid-catalyzed oxidation of glucose

Novel pathway using naturally occurring enzymes (*Bioprospecting*)



Co-expression of 3 genes in E. coli



Build-up of 1st intermediate indicates a limitation with the 2nd enzyme (MIOX)

Moon et al, 2009. Appl. Environ. Microbiol. 75(3):589-595

A closer look at MIOX...



→ High MI production by 1st enzyme (Ino1) is desired.
→ Easier said than done...

w/o MI

Moon et al, 2009. Appl. Environ. Microbiol. 75(3):589-595

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Enzyme Co-localization

(Collaboration with Dr. John Dueber, SynBERC*)



MI = myo-Inositol

* QB3 Fellow, University of California, Berkeley, USA Synthetic Biology Engineering Research Center (SynBERC)

Effect of Co-Recruitment on Enzyme Activity

Fixed enzyme induction level, variable scaffold induction levels



Moon et al, 2010. Metabolic Engineering, 12, 298-305

Manipulating Glucose Metabolism



Modulation of glucokinase (glk) may be used to redirect glucose and increase pathway productivity with glucose as sole carbon source

Glk Expression Library



Anderson Promoter Library, http://partsregistry.org

Glk Expression Library



On glucose, growth rate varies with Glk activity

Solomon et al, 2013. ACS Synthetic Biology, 2:126-135.

Gluconate Productivity as a Function of Glk Activity



Reductions in Glk activity lead to increases in molar yield but only when endogenous needs are met

Solomon et al, 2013. ACS Synthetic Biology, 2:126-135.

Inverters as a glucose valve



Biological inverters show a dose dependent response

Solomon et al, 2012. *Metabolic Engineering*, 14:661-671.

Inverter Effect on Productivity



Solomon et al, 2012. Metabolic Engineering, 14:661-671.

Effect of Timing of Induction



Summary of Glucaric Acid Production

- Bioprospecting for Part Selection resulted in identification of enzymes necessary to create a novel pathway.
- Use of Synthetic Biology Devices led to increases in productivity.
- Host engineering may provide a means to further improve flux and titers.

3-Hydroxybutyrolactone (3-HBL)

- Key Intermediate in Higher, Chiral Synthesis of Solvents (e.g. Furan Derivatives) and Pharmaceuticals (e.g. Statins)
- Wholesale Cost ~ \$450/kg (\$20-50/gram for lab-scale quantities)
- No Known Biological Routes towards DHBA or 3-HBL.







The 3-Hydroxybutyrate Pathway



Enzyme	Organism of Origin	Properties	Reference	
bktB	R. eutropha H16	Acetyltransferase, broad substrate range (C_4-C_6)	Slater, 1998	
hbd	C. acetobutylicum 824	Dehydrogenase, forms S stereoisomer product	Boynton, 1996	
phaA	R. eutropha H16	Acetyltransferase, used in biopolymer synthesis	Schubert, 1988	
phaB	R. eutropha H16	Dehydrogenase, forms R stereoisomer product	Schubert, 1988	
tesB	<i>E. coli</i> K12 Thioesterase, Very broad substrate range (C_4-C_{16})		Huisman, 1991	
thil	C. acetobutylicum 824	Acetyltransferase, high activity	Stim-Herndon, 1995	

Tseng et al, 2009. Appl. Environ. Microbiol. 75(10):3137-3145. >2 g/L R- or S- 3HB

3-Hydroxyvalerate Synthesis



CoA-activation Enzymes



Chiral 3HV Production

✓ Activation of propionate is crucial

✓Activation mechanism determines the product distribution



Tseng et al, 2010, Microbial Cell Factories. 9:96

From 3HB/3HV to DHBA?

3-hydroxybutyrate & 3-hydroxyvalerate biosynthetic pathway





Production of 3,4-Dihydroxybutyrate (DHBA) Glucose + Glycolate



All cultures were *E. coli* MG1655(DE3) *end*^{A⁻} rec^{A⁻} grown in LB for 72 hours and were supplied with glycolate. Martin et al, 2013. *Nat. Commun.* 4, 1414

Direct Synthesis of Glycolate, DHBA and 3-HBL from Glucose



Effect of Glucose Concentration on DHBA



DHBA

3HBL

Glucose Feed	Molar Yield on Glucose	% of Theoretical Yield	([3,4- DHBA]+[3HBL])
			[3HB]
0.8%	0.095	14.4	3.514
1.0%	0.141	21.3	2.557
1.5%	0.136	20.6	2.242

An Unexpected Product



Extending the Hydroxyacid Pathway



Summary of Hydroxyacids Production

- Established a versatile platform for biological synthesis of chiral hydroxyacids.
- Demonstrated 1st pathway for biological production of 3-hydroxybutyrolactone from simple (and sole) carbohydrate substrates.
- Reliance on promiscuity of enzymes results in wide range of productivities for novel substrates.
 - Bioprospecting and Protein Engineering

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Current Group

Dr. Christopher Reisch Joana Rodrigues **Micah Sheppard Eric Shiue** Sue Zanne Tan

Former Students

Dr. Diana Bower

Dr. Collin Martin

- Dr. Tae Seok Moon
- Dr. Kevin Solomon
- Dr. Hsien-Chung Tseng

Former Post-Docs

Dr. Effendi Leonard Dr. David Nielsen Dr. Sang-Hwal Yoon

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